



Original Article

A survey of equine anti-*Listeria monocytogenes* antibodies using Latex Agglutination Test in southeast of Iran

Poona Faramarzpour¹, Ehsanollah Sakhaee*², Mehdi Golchin³, Balal Sadeghi⁴

¹ School of Veterinary Medicine graduate, Shahid Bahonar University of Kerman, Kerman, Iran

² Department of Clinical Sciences, School of Veterinary Medicine, Shahid Bahonar University of Kerman, Kerman, Iran

³ Department of Pathobiology, School of Veterinary Medicine, Shahid Bahonar University of Kerman, Kerman, Iran

⁴ Department of Food Hygiene and Public Health, School of Veterinary Medicine, Shahid Bahonar University of Kerman, Kerman, Iran

ARTICLE INFO

Received: 3 August 2017

Accepted: 1 October 2017

KEY WORDS :

Listeria monocytogenes
Latex Agglutination Test
Horse
Iran

ABSTRACT

Listeriosis is a zoonotic disease in humans and a wide range of domestic and wild animals and also some birds. The main purpose of the current study is to determination of anti-*Listeria monocytogenes* sero-prevalence in horses by latex agglutination test in southeast of Iran. A total of 163 serum samples were obtained from apparently healthy horses of equestrian clubs in Kerman and Yazd provinces - Iran. *Listeria monocytogenes* antibodies were found in 34 out of the 163 sera (20.85%). The latex agglutination test can be considered as an appropriate screening test in the early stages of diagnosis.

ردیابی پادتن ضد لیستریا مونوسیتوژنز با روش لاتکس اگلوتیناسیون در اسبان جنوب شرق ایران

پونا فرامرزپور^۱، احسان اله سخائی*^۲، مهدی گلچین^۳، بلال صادقی^۴

^۱ گروه علوم درمانگاهی دانشکده دامپزشکی دانشگاه شهید باهنر کرمان، کرمان، ایران

^۲ گروه علوم درمانگاهی دانشکده دامپزشکی دانشگاه شهید باهنر کرمان، کرمان، ایران

^۳ گروه پاتوبیولوژی دانشکده دامپزشکی دانشگاه شهید باهنر کرمان، کرمان، ایران

^۴ گروه بهداشت مواد غذایی دانشکده دامپزشکی دانشگاه شهید باهنر کرمان، کرمان، ایران

چکیده

لیستریوز یک بیماری مشترک میان انسان و طیف وسیعی از دامهای اهلی، وحشی و پرندگان می باشد. هدف از انجام مطالعه ی حاضر ردیابی پادتن ضد لیستریا مونوسیتوژنز با روش لاتکس اگلوتیناسیون در اسبان جنوب شرق ایران می باشد. برای این منظور تعداد ۱۶۳ نمونه سرم از اسب های به ظاهر سالم باشگاه های سوارکاری استان های یزد و کرمان تهیه شد. نتایج مطالعه حاکی از حضور پادتن در ۳۴ نمونه از مجموع ۱۶۳ سرم مورد بررسی (۲۰/۸۵ درصد) می باشد. بر این اساس به نظر می رسد که لاتکس اگلوتیناسیون آزمایش مناسبی برای غربالگری اولیه به منظور تشخیص بیماری می باشد.

واژه های کلیدی: لیستریا مونوسیتوژنز، آزمایش لاتکس اگلوتیناسیون، اسب، ایران

*Corresponding author: Ehsan_Sakhaee@uk.ac.ir

Introduction

Listeriosis is one of the food-borne zoonosis diseases around the world caused by *Listeria* species. This causes considerable morbidity and mortality in humans and animals and the bacterium is considered an important food borne pathogen [1]. *Listeria monocytogenes* is ubiquitous, aerobic, gram positive and rod-shape bacterium, which is capable of causing severe disease in many species as sheep, cattle, goats, horse, humans and chickens [7,19]. There is six species of listeria in this genus that categorized into three groups as genome analyze, that first group contains *L. monocytogenes*, *L. innocua* and *L. welshimeri*, the second one *L. ivanovii*, and *L. seeligeri*, and the third include *L. grayi* [8]. Listeriosis occurs as a sporadic disease in horses with highly pathogenic intracellular, non-acid resistant bacteria and is clinically characterized by meningo-encephalitis. Signs associated with the nervous syndrome include paralysis of the mandibular and pharyngeal muscles, difficulty walking, inappetence, polydipsia, loss of body weight, and collapse. Following the first observation of signs, animals usually die within 3-10 days [9,11]

Materials and Methods

Sample collection and processing

A total of 163 serum samples were collected from 93 male and 70 female (clinically healthy) at 9 equestrian clubs in Kerman and Yazd provinces, Iran. The breed composition comprised of Arab and Arab cross horses (50 %) and non-Arab horses (50%) including Darreh-Shouri (5%), Turkmen (12 %), Thoroughbred (3 %) and mixed breeds (30 %).

After collection, the samples were submitted to Microbiology laboratory of School of Veterinary Medicine, Shahid Bahonar University of Kerman, Iran. Sera were stored at -20°C until analysis.

Latex Agglutination Test (LAT)

Latex Agglutination Test Kit (Zist Faravard Pars Co, Rasht, Iran) was used according to the manufacture's protocol. Briefly, 10 μl of serum and 10 μl of antigen coated latex particles were added to on an agglutination card and mixed with a plastic stirrer. The card was rocked from side to side for up to 5 min to provoke the agglutination reaction. Specimens that showed agglutination during this period were recorded as positive, and otherwise negative. Incomplete agglutination recorded as suspected. Specificity and sensitivity assurance was carried out by testing positive and negative controls on daily basis.

Results

According to the Table 1 and 2, antibodies were detected in 34 sera (20.85%) out of 163 samples (21 cases ≤ 7 years old and 13 cases > 7 years old). Based on the results presented on Table 1 and 2, 21 (15 cases ≤ 7 years old and 6 cases > 7 years old) male (12.88%) and 13 (6 cases ≤ 7 years old and 7 cases > 7 years old) mares (7.98%) were sero-positive against listeriosis.

Table 1. Number and frequency (%) of positive, suspected and negative samples among 163 equine serum samples

Gender	Positive		Suspect		Negative		Total
	No*	F (%) **	No	F (%)	No	F (%)	No
Stallion	21	12.88	22	13.50	50	30.67	93
Mare	13	7.98	22	13.50	35	21.47	70
Total	34	20.86	44	27	85	52.14	163

*: Number of cases ** : Frequency (percent)

Table2. The number and frequency of positive, negative and suspected samples (based on the presence of antibodies against *Listeria monocytogenes*) out of 163 horses in Kerman and Yazd provinces, Iran

Province	Gender	Age								Total												
		≤ 7 years old				> 7 years old				P	N	S	T									
		P	N	S	T	P	N	S	T													
		No	F	No	F	No	F	No	F	No	F	No	F	No	F							
Yazd	Stallion	7	14.58	31	64.58	10	20.83	48	14.28	1	14.28	5	71.42	7	8	14.54	32	58.19	15	27.27	55	27.27
	Mare	3	8.57	20	57.14	12	34.28	35	8.33	4	33.33	7	58.33	12	4	8.52	34	51.06	19	40.42	47	40.42
Kerman	Stallion	8	36.36	11	50	3	13.63	22	31.25	5	43.75	4	25	16	13	34.21	18	47.36	7	18.42	38	18.42
	Mare	3	27.27	6	54.54	2	18.18	11	50	6	41.66	1	8.33	12	9	39.13	11	47.82	3	13.04	23	13.04
	Total	21	18.10	68	58.62	27	23.27	116	27.65	13	36.17	17	36.17	47	34	20.85	85	52.14	44	26.99	163	26.99

Discussion

Our results showed the total seropositivity for all 163 horses examined in this study was 20.85%. *Listeria monocytogenes* widely exists in the water, soil, plants, feces and feedstuff such as silage, vegetables and moldy forage. This bacteria is considered as one of the most

important sources of infection in both domestic and wild animals and also some birds [16]. The prevalence of listeriosis has been mostly reported during the winter which can be due the fact that silage is frequently fed to animals in this season and also the pregnancy of animals predisposes them to this

infectious disease [12]. Listeriosis in farm animals often occurs in three main forms: encephalitis, septicemia and abortion [5]. However, the encephalitis form has a lower occurrence rate in horses [5,7]. Teruya *et al.*, (1977) conducted a study on equine listeriosis in Brazil using tube agglutination method and reported that 22.7% of 838 examined horses were seropositive. Solmaz *et al.*, (2002) carried out a similar study in Turkey using the same method and found 176 positive (86.7%) out of 203 horses [15,17]. Guclu *et al.*, (2007) showed 62 cases among 100 tested horses were sero-positive against *L. monocytogenes* using the Osebold absorption test. Anti *L. monocytogenes* antibodies were detected at 1:100, 1:200 and 1:400 titers in 29 (46.7%), 31 (50%) and 2 (3.2%) animals, respectively [20]. The results of the present study are similar to the study conducted by Teruya *et al.*, (1977) and Guclu *et al.*, (2007). However, higher prevalence in the study conducted by Solmaz *et al.*, can be due to factors such as climate variation, feedstuff and health management of the animals.

The results of the current study showed more sero-positivity among older males in contrast with previous study showed there is no relationship between age and sex and incidence of listeriosis. Saqib *et al.*, (2015) reported that 23.5% out of 183 equine serum samples were seropositive and number of seropositive males was higher than females [13]. Results of the present study in Yazd and Kerman provinces showed that the higher anti *L. monocytogenes* antibody titer found in horses in Kerman is possibly due to moderate climate and also the lower temperature and different management and any other factors or

any combination(s). The obtained results of the separate studies in each province showed that the sero-positivity in the old males of Kerman follows the general rule which was mentioned earlier. The results of serological studies have shown that the presence of anti *L. monocytogenes* antibody may be widespread worldwide, but the absence of definitive clinical diagnosis and data makes it impossible to estimate the true incidence of listeriosis in animals and humans. These results have also revealed that the seroprevalence of positive antibody titer against *Listeria monocytogenes* in animals varies widely with species tested, geographic location, season, assay type and the criteria used to define positive results [2,3,4,9,18]

It has been reported that the antigenic relationship between various serotypes of *L. monocytogenes* and a number of gram-positive and gram-negative bacteria (such as *Staphylococcus aureus*, *Streptococcus faecalis*, *Arcanobacter pyogenes*, *Bacillus subtilis* and *Escherichia coli*) may cause false-positive results in serological tests [10,14]. The latex agglutination test can be considered as an appropriate screening test in the early stages of diagnosis, because mentioned method has a favorable sensitivity and specificity. It is also a fast and cheap method which can be performed in all laboratories.

Conclusion

The current study detected anti *L. monocytogenes* antibody in the horses' population of Kerman and Yazd provinces, Iran. Mentioned results showed that the higher antibody titer found in Kerman province is possibly due to moderate climate and also the

lower temperature and different management and any other factors or any combination(s).

References

- [1] Voelter-Ratson K, Pot S, Florin M, Spiess B. Equine keratomycosis in Switzerland: a retrospective evaluation of 35 horses (January 2000–August 2011). *Equine veterinary journal*. 2013;45(5):608-12.
- [2] Brooks D, Andrew S, Dillavou C, Ellis G, Kubilis P. Antimicrobial susceptibility patterns of fungi isolated from horses with ulcerative keratomycosis. *American journal of veterinary research*. 1998;59(2):138-42.
- [3] Gemensky-Metzler AJ, Wilkie DA, Kowalski JJ, Schmall LM, Willis AM, Yamagata M. Changes in bacterial and fungal ocular flora of clinically normal horses following experimental application of topical antimicrobial or antimicrobial-corticosteroid ophthalmic preparations. *American journal of veterinary research*. 2005;66(5):800-11.
- [4] Pearce JW, Giuliano EA, Moore CP. In vitro susceptibility patterns of *Aspergillus* and *Fusarium* species isolated from equine ulcerative keratomycosis cases in the midwestern and southern United States with inclusion of the new antifungal agent voriconazole. *Veterinary ophthalmology*. 2009;12(5):318-24.
- [5] Gaarder J, Rebhun W, Ball M, Patten V, Shin S, Erb H. Clinical appearances, healing patterns, risk factors, and outcomes of horses with fungal keratitis: 53 cases (1978-1996). *Journal of the American Veterinary Medical Association*. 1998;213(1):105-12.
- [6] Reed Z, Thomasy S, Good K, Maggs D, Magdesian K, Pusterla N, et al. Equine keratomycoses in California from 1987 to 2010 (47 cases). *Equine veterinary journal*. 2013;45(3):361-6.
- [7] Matthews A. The aetiopathogenesis of infectious keratitis in the horse. *Equine veterinary journal*. 1994;26(6):432-3.
- [8] Moore C, Heller N, Majors L, Whitley R, Burgess E, Weber J. Prevalence of ocular microorganisms in hospitalized and stabled horses. *American journal of veterinary research*. 1988;49(6):773-7.
- [9] McLaughlin S, Brightman A, Helper L, Manning J, Tomes J. Pathogenic bacteria and fungi associated with extraocular disease in the horse. *Journal of the American Veterinary Medical Association*. 1983;182(3):241-2.
- [10] Nardoni S, Sgorbini M, Barsotti G, Corazza M, Mancianti F. Conjunctival fungal flora in healthy donkeys. *Veterinary ophthalmology*. 2007;10(4):207-10.
- [11] Sgorbini M, Barsotti G, Nardoni S, Mancianti F, Rossi S, Corazza M. Fungal flora of normal eyes in healthy newborn foals living in the same stud farm in Italy. *Journal of equine veterinary science*. 2008;28(9):540-3.
- [12] Markey B, Leonard F, Archambault M, Cullinane A, Maguire D. *Clinical Veterinary Microbiology E-Book*: Elsevier Health Sciences; 2013.
- [13] Johns IC, Baxter K, Booter H, Hicks C, Menzies-Gow N. Conjunctival bacterial and fungal flora in healthy horses in the

Acknowledgments

This research was financially supported by the Research Council of Shahid Bahonar University of Kerman, Iran.

Conflict of Interest

There is no conflict of interest.

- UK. *Veterinary ophthalmology*. 2011;14(3):195-9.
- [14] Rosa M, Cardozo LM, da Silva Pereira J, Brooks DE, Martins ALB, Florido PSS, et al. Fungal flora of normal eyes of healthy horses from the State of Rio de Janeiro, Brazil. *Veterinary ophthalmology*. 2003;6(1):51-5.
- [15] Whitley R, BURGESS EC, Moore C. Microbial isolates of the normal equine eye. *Equine Veterinary Journal*. 1983;15(S2):138-40.
- [16] Andrew SE, Nguyen A, Jones GL, Brooks DE. Seasonal effects on the aerobic bacterial and fungal conjunctival flora of normal thoroughbred brood mares in Florida. *Veterinary ophthalmology*. 2003;6(1):45-50.
- [17] Araghi-Sooreh A, Navidi M, Razi M. Conjunctival bacterial and fungal isolates in clinically healthy working horses in Iran. *Kafkas Univ Vet Fak Derg*. 2014;20:625-7.
- [18] Pisani EHR, de Moraes Barros PS, de Avila FA. Microbiota conjuntival normal de equinos Departamento de. *Brazilian Journal of Veterinary Research and Animal Science*. 1997;34(5):261-5.
- [19] Andrew S, Brooks D, Smith P, Gelatt K, Chmielewski N, Whittaker C. Equine ulcerative keratomycosis: visual outcome and ocular survival in 39 cases (1987–1996). *Equine Veterinary Journal*. 1998;30(2):109-16.
- [20] Ledbetter EC, Patten VH, Scarlett JM, Vermeylen FM. In vitro susceptibility patterns of fungi associated with keratomycosis in horses of the northeastern United States: 68 cases (1987–2006). *Journal of the American Veterinary Medical Association*. 2007;231(7):1086-91.
- [21] Galán A, Martín-Suárez E, Gallardo J, Molleda J. Clinical findings and progression of 10 cases of equine ulcerative keratomycosis (2004–2007). *Equine Veterinary Education*. 2009;21(5):236-42.
- [22] Voelter-Ratson K, Monod M, Unger L, Spiess BM, Pot SA. Evaluation of the conjunctival fungal flora and its susceptibility to antifungal agents in healthy horses in Switzerland. *Veterinary ophthalmology*. 2014;17(s1):31-6.
- [23] Coad C, Robinson N, Wilhelmus K. Antifungal sensitivity testing for equine keratomycosis. *American journal of veterinary research*. 1985;46(3):676-8.
- [24] Verneuil M, Durand B, Marcon C, Guillot J. Conjunctival and cutaneous fungal flora in clinically normal dogs in southern France. *Journal de Mycologie Médicale/Journal of Medical Mycology*. 2014;24(1):25-8.
- [25] Brooks D. Equine keratomycosis: an international problem. *Equine Veterinary Education*. 2009;21(5):243-6.
- [26] Liu J, Li J, Huo J, Xie H. Identification and quantitation of conjunctival aerobic bacterial flora from healthy residents at different ages in Southwest China. *African Journal of Microbiology Research*. 2011;5(3):192-7.
- [27] Gionfriddo J, Rosenbusch R, Kinyon J, Betts D, Smith T. Bacterial and mycoplasmal flora of the healthy camelid conjunctival sac. *American journal of veterinary research*. 1991;52(7):1061-4.
- [28] Araghi-Sooreh A, Mokhber-Dezfuli M, Mohammadi-Chorsi M. Identification of fungal isolates from conjunctival sac in healthy goats. *Journal of Veterinary Research*. 2013;68(4):327-32.
- [29] Davidson H, Rogers D, Yearly T, Stone G, Schoneweis D, Chengappa M. Conjunctival microbial flora of clinically normal pigs. *American journal of veterinary research*. 1994;55(7):949-51.