



Cell suspension culture of yew: an alternative method of taxol production

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Abstract

Taxol is one of the most important anti-cancer drugs in the world, which is obtained from the genus of the *Taxus*. Taxol, with its antimicrobial properties, causes the death of proliferating cells by preventing the formation of abnormally dividing spindles. Abnormal division stops DNA transcription in the G2 / M division of mitosis and thus causes the proliferation of proliferating cells. Due to the low amount of taxol in the tissues of yew and also the very low growth of this plant, meeting the therapeutic need for this drug is the most important issue facing scientists. Commercially, taxol synthesis seems unlikely due to limited resources and the possibility of over-consumption of a natural product that could be found in endangered yew species. Therefore, a suitable alternative method for taxol production should be used. Yew cell suspension culture is one of the most important alternatives for long-term and sustainable production of taxol. This review is about cell suspension culture in different yew species.

Keywords: taxol, anti-cancer drug, secondary metabolite, cell suspension culture, taxus

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Introduction

Taxol is a very important secondary metabolite that has a diterpenoid structure (Jordon and Wilson, 1995). This valuable substance called generic name paclitaxel is the most effective anti-cancer drug of plant origin (Liu et al., 2016). Today, cancer is one of the deadliest diseases that human beings have ever struggled with and is one of the most common causes of death in the world (Shoeb, 2006). Uterine, breast and ovarian cancers are among the most common cancers among

Plants have been used as a source of natural compounds agents to cancer with minimal side effects. Today, chemotherapy is the most important and main method of cancer treatment. In this method, anti-cancer drugs are used alone or in combination with other drugs (Siegel and Jemal, 2012). During the 1960s, the National Cancer Institute (NCI) began research into the anticancer effects of medicinal Plants and eventually discovered the anticancer drug taxol (Itokawa and Lee, 2002). Clinical use of Taxol was approved by the FDA after a series of clinical trials in 1983-86 (Wall, 1993) and has been used as the most effective drug for the treatment of breast and ovarian cancer and for the treatment of other cancers, including cancers Lung, bladder,

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women worldwide. In recent decades, medicinal



Fig. 1. Taxus trees

intestines and stomach are used (Holmes et al., 1995. Suffness and Wall, 1995).

Taxol function

Taxol polymerizes tubulin and stops it from depolymerizing, thus stopping the mitosis division cycle (Hortuwitz, 1992). Taxol stops the transcription of DNA in the G2-M stage by forming an abnormal division spindle, causing cell death and preventing cell proliferation (Jennewein and Croteau, 2001). Taxol is a trademark of Paclitaxel, registered by Bristol-Myers Squibb. All taxus species contain the valuable substance taxol, including *T. baccata*, *T. brevifolia*, *T. cuspidata*, *T. Canadensis*, *T. chinensis*, *T. Canadensis*, *T. yunnanensis* and *Taxus x media*, *Taxus x hunnewelliana* (Cope, 1998). Taxol was first found in the skin, roots and other parts of *T. brevifolia* in 1971 (Wani et al., 1971).

Distribution

Yew (taxus) is a tree of the genus *Taxus*, order of the Taxales and the family of Taxaceae, which is distributed in temperate regions, especially in the Northern Hemisphere, Europe, Asia and North America (Bedi, 1996). Eight species of yew have been reported and so far 72 varieties have been introduced for yew, the difference of which is based on height, leaf colour and cold resistance (Itokawa and Lee, 2003). Fossil studies show that yew trees are 190 million years old and the oldest fossils belong to the Miocene and Pliocene periods. In later periods, yew trees were seen with beech and hornbeam trees (Mossadegh, 1993). Yew is a shade-loving tree and is resistant to low light environments and can perform its

physiological activities even in low light. Yew leaves are narrow and long and are dark green and clear on the upper surface of the leaflet, but light green on the lower surface. The leaves are 2-3 cm long and 2-3 mm wide. The flowers of the plant are male and female, located on two separate bases (Mossadegh, 1993). Yew is a slow-growing but long-lived tree that has been observed in nature for 1000 to 1500 years (Fig 1).

Taxol

Taxol is a type of alkaloid diterpene that is obtained from all different species of yew and is very important as an anti-cancer drug (Mihaljevic and Bjedow, 2002). Taxol with the scientific name of paclitaxel has a chemical formula of $C_{47}H_{51}NO_{14}$ and a molecular weight of 853.92 dalton, which has a melting point of 158-160 °C (Jennewein and Croteau, 2001). All Taxanes are derivatives of diterpenoids, of which more than 350 kind have been identified in various yew species, the most important of which are taxol, 10-deacetylbaccatin III, baccatin III, and Taxinine. In general, the taxol synthesis pathway consists of nineteen enzymatic steps (Croteau et al., 2006). The first step in the reaction is to ringing of the geranylgeranyl diphosphate (GGPP) compound (Hezari et al., 1995).

Then, during a series of hydroxylation (Croteau et al., 2006) and acetylation (Walker and Croteau, 2000) reactions, very important intermediates of 10-deacetylbaccatin III and baccatin III are formed. After the formation of baccatin II, the α -Phenylalanine side chain is attached to it (Walker et al., 2004), and after several stages of enzymatic reactions (Onrubia et

al., 2013, Walker et al., 2002), the valuable substance of taxol is finally synthesized (Walker et al., 2002) (Fig II).

But the main problem is the small amount of taxol in the bark of the yew tree, the amount of taxol varies from 0.001 to 0.05 in different species of yew (Schippmann, 2001). For this reason, many attempts were made to replace other methods in

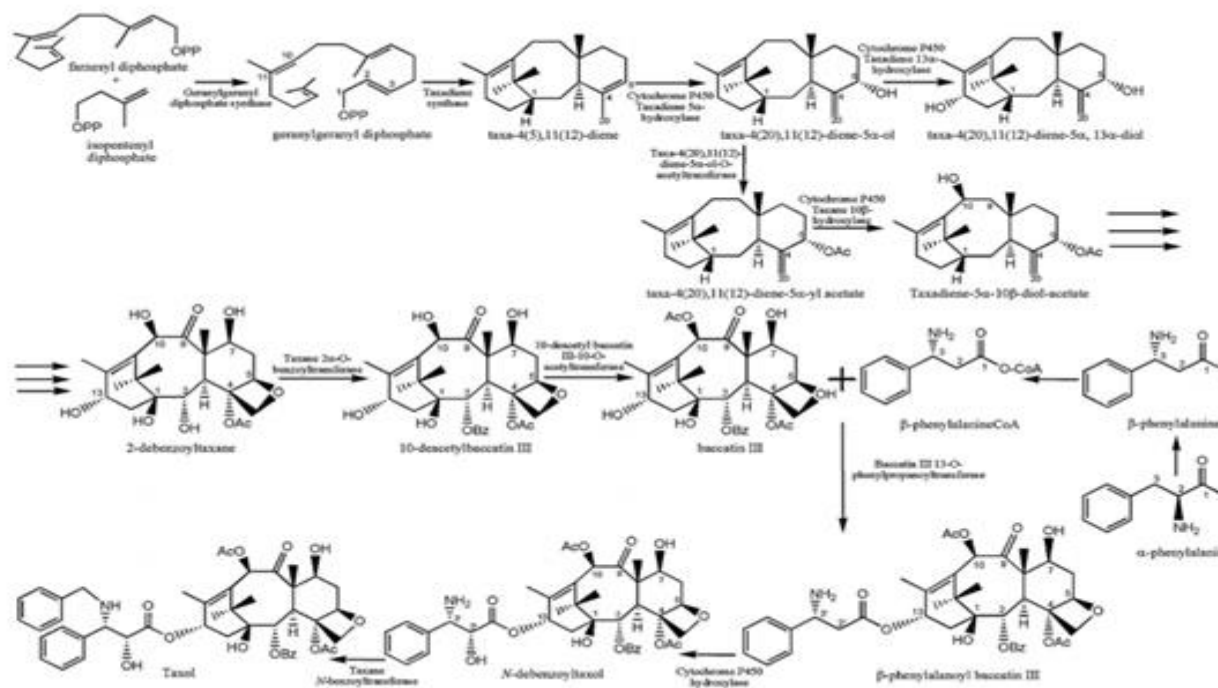


Fig. II. Biosynthetic path to Taxol

About 300 mg of taxol is obtained from each adult yew plant, while each patient needs 2.5 to 3 grams of medicine for treatment. Almost every cancer patient needs 6 yew trees during their treatment. Due to the increase in the number of patients and the lack of natural resources of the yew plant, the production of taxol from the plant as the only source of this valuable drug is not economical because to get one kilogram of taxol from the plant requires cutting down 2,000 - 2,500 of 200-year-old yew trees (Liu, 2016, Malik et al., 2011, Croteau et al., 2006, Onrubia et al., 2013). The production of taxol from this natural source causes indiscriminate cutting and extinction of yew trees. Over the past decade, researchers have been looking for new ways to improve production conditions and reduce the price of this valuable drug to meet the needs of cancer patients and specialist clinics (Jennewein and Croteau, 2001).

the production of taxol. Other alternative methods, such as the semi-synthetic reaction of precursors such as baccatin III and 10-deacetyl baccatin III, which are present in larger quantities in the plant, as well as its production by biotechnological methods were considered (Stierle et al., 1995). In 1994, the first success in the production of taxol by chemical synthesis was achieved (Nicolau et al., 1994). However, the complexity of the taxol biosynthesis pathway, low yield and limited ability to perform chemical synthesis led to the consideration of an alternative method for the supply of this compound. Semi-synthetic taxol systems are one of these methods, which is done due to the precursors of baccatin and 10 baccatin in yew leaves. During the last two decades, biotechnology methods, especially yew cell culture, have been used in the production of taxol, which is one of the sustainable production

methods of this compound due to its independence from the environment (Zhong, 2002, Tabata, 2004).

Cell suspension culture

The most common plant cell culture systems are callus cultures, which are dedifferentiated cells. In a cell culture system, cells grow in a liquid culture medium and are constantly shaken. Cell culture should be subcultured periodically, which depends on the speed of cell growth (Mustafa et al., 2011). Continuous harvesting of the plant to produce taxols may not be a sufficient source to meet the clinical demands. so, interest in taxol production increased through culture of cell suspension. Cell cultures have a faster growth rate than whole plant cultures and show an increase in production compared to whole plants against the addition of treatments, which results in increased production in less time (Fett-Neto et al., 2004, Gandhi et al., 2015, Rao and Ravishankar, 2002). Due to the totipotency property in plants, it can be expected that all the compounds in the whole plant will be observed in its cell culture (Rao and Ravishankar, 2002). In cell cultures, it should be noted that the production of the active substance is influenced by two factors: production per cell and cell mass size, which in most cases, the optimal conditions for both factors are in conflict with the other to solve this problem. Two-stage culture is suggested that in the first stage, growth conditions are in favor of cell growth, and after obtaining the appropriate cell mass, the second stage begins, which is in favor of the production of the desired metabolite (Rao and Ravishankar, 2002, Zhong, 2002, Malik et al., 2011). Accumulation of drugs in large-scale suspension culture is considered as the best, sustainable and long-term alternative for taxol production (Lee et al., 2010). Many scientists prefer two-phase culture with the use of elicitor to increase productivity and maintain cell integrity.

Use of elicitors

Elicitors are compounds of biological and non-biological origin that stimulate secondary metabolites (Dornenburg and Knorr, 1995). Elicitors are biological stress agents and their use in low concentrations on the plant induces a stress

response (real stress or good stress) leading to the production of secondary metabolites in plants (phenolic compounds, flavonoids, alkaloids and ...) And also protects the plant against living and non-living factors without affecting the efficiency of the plant (Vazquez-Hernandez et al., 2019, Vargas-Hernandez et al., 2017, Cardenas-Manríquez et al., 2016). Elicitors are used to stimulate the production of plant metabolites (Mejía-Teniente et al., 2010). Researchers focus on the effects of an elicitor. Methyl jasmonate is an elicitor that researchers have studied extensively on its effect on taxol biosynthesis (Ketchum et al., 1999, Bonfill et al., 2006). So far, many different compounds have been used to induce taxol in cell culture. Improvement of taxol production in culture of *T. wallichiana* Zucc cell suspension has been reported with the addition of IAA-conjugates (IAA-glycine, IAA-phenylalanine, IAA-alanine and IAA-aspartic acid) (Jha et al., 1998). Phenylalanine and vanadyl sulfate were used in *Taxus baccata* L. cells (Brincat et al., 2002). A study by Zhong found that L-phenylalanine, as a precursor to the taxol side chain, had a significant effect on increasing taxol synthesis in the suspension culture (Zhang et al., 2002). Also, application of methyl jasmonate to *T. cuspidate* caused a different increase in the expression of genes in the taxol production pathway, which showed that the transcripts of the genes at the end of the pathway are much less than the transcripts of the genes at the beginning of the pathway. The importance of these genes was in metabolite engineering (Nims et al., 2006). Study on ethylene inhibitors such as silver nitrate and cobalt chloride, along with their combination in *Taxus spp.* suspension culture showed an increase in taxol production (Zhang and Wu, 2003). The use of three elicitors of chitosan, methyl jasmonate and silver on *T. chinensis* culture showed that the combined use of these three elicitors increased taxol to forty times control, ten times the use of silver alone, six times the use of chitosan alone and twice the use of only methyl jasmonate, which has been argued that different elicitor induce different defense responses (Zhang et al., 2000, Sabater-Jara and Pedreño, 2013). Many factors affect the production of taxol, including: Inducing factors, addition time, culture days and culture stage,

different concentrations of elicitors have been shown to affect cell culture and taxol production (Linden and Phisalaphong, 2000). Khosroshahi et al (2006) tried to improve the production of taxol by combining different induction factors in the suspension culture of *Taxus baccata*. They developed suspension culture using Gamborg's B5 medium with various inducers such as vanadyl sulfate, cobalt chloride, silver nitrate ammonium citrate, phenylalanine, methyl jasmonate, salicylic acid and fungal elicitor (Khosroshahi et al., 2006). In another study, the ability of dimethylsulfoxide (DMSO) to induce the synthesis and release of taxanes in the cell suspension culture of *Taxus baccata* was investigated and showed that addition of 5% DMSO had the best performance of taxanes in both sections (Kajani et al., 2012).

Production of taxol in bioreactors

With the advancement of biotechnology, semi- or total synthesis of taxol has been done by culturing endophytes and in cell suspension culture. Taxol production in bioreactors is continuous and abundant and is also considered as an excellent alternative method for long-term production of taxol without destroying the plant in its natural habitat (Lee et al., 2010). Several types of bioreactors are designed to plant cells culture in

large volumes (Roberts and Shuler, 1997). In a study, Stirred, Airlift and Wave bioreactors were used for the production of taxol and baccatin III from cultured cells with calcium alginate. The highest amount of taxol and baccatin III produced was related to Stirred bioreactor, followed by Wave and Airlift bioreactors (Bentebibel et al., 2005). Today, for commercial production of Taxol, bioreactors with a volume of more than 75,000 liters are used by Phyton Biotech, ESCA genetic, Samyang Genex, Nattermann (Germany) (Frense, 2007. Kolewe et al., 2008).

Metabolic engineering

Simulation of the taxol-producing gene has also been conceived but is still ongoing. Large-scale biological process engineering of plant cells is a cost-effective method and the responsiveness of various factors studied has further highlighted the application of this method. Endophytic fungi of the species *Taxus* have also been extensively studied for taxol production, however, the stable production of these microorganisms has not yet been proven and also the purification of intact tissues compared to plant cell culture is probably long and it is expensive (Cusido et al., 2002).

References

- Brincat, M. C., D. M. Gibson and M. L. Shuler,** 2002. 'Alterations in Taxol production in plant cell culture via manipulation of the phenylalanine ammonia lyase pathway'. *Biotechnology Progress*, 18, 1149-56.
- Bedi, Y., R. Ogra, K. Koul, B. Kaul and R. Kapil,** 1996. 'Yew (*Taxus* spp.) A new look on utilization, cultivation and conservation'. Supplement to cultivation and utilization of medicinal plants. *Jammu-Tawi: Regional Research Laboratory*.
- Bonfill, M., O. Expósito, E. Moyano, R.M. Cusido, J. Palazón and M.T. Piñol,** 2006. 'Manipulation by culture mixing and elicitation of paclitaxel and baccatin III production in *Taxus*'. *In vitro Cellular and Developmental Biology – Plant*, 42, 422-426.
- Bentebibel, S, E. Moyano, J. Palazon, RM. Cusido, M. Bonfill, R. Eibl,** 2005. 'Effects of immobilization by entrapment in alginate and scale-up on paclitaxel and baccatin III production in cell suspension cultures of *Taxus baccata*'. *Biotechnol Bioeng* 89:647-55.
- Cusido, R. M. Palazon, J. Bonfill, M. Navia-Osorio, A. Morales, C. and Pinol, M. T.** 2002. 'Improved paclitaxel and baccatin III production in suspension cultures of *Taxus media*'. *Biotechnology Progress*, 18, 418-23.
- Cope, EA,** 1998. 'Taxaceae: the genera and cultivated species'. *Bot Rev.* 64(4):291-322.
- Croteau, R., REB. Ketchum, RM. Long, R. Kaspera and M. Wildung,** 2006. 'Taxol biosynthesis and molecular genetics'. *Phytochemistry Reviews* 5: 75-97.

- Croteau, R., REB. Ketchum, RM. Long, R. Kaspera and M. Wildung**, 2006. 'Taxol biosynthesis and molecular genetics'. *Phytochemistry Reviews* 5: 75-97.
- Cardenas-Manríquez, G., I. Vega-Muñoz, A. Villagómez-Aranda, M. León-Galvan, A. CruzHernandez, I. Torres-Pacheco, R. Rangel-Cano, R. Rivera-Bustamante and R. GuevaraGonzalez**, 2016. 'Proteomic and metabolomic profiles in transgenic tobacco (*N. tabacum xanthi* nc) to CchGLP from *Capsicum chinense* BG-3821 resistant to biotic and abiotic stresses'. *Environmental and Experimental Botany*. 130, 33–41.
- Dornenburg, H. and D. Knorr**, 1995. 'Strategies for the improvement of secondary metabolite production in plant cell cultures'. *Enzyme and Microbial Technology*, 17, 674-684.
- Fett-Neto, AG., H. Aoyagi, H. Tanaka and F. Dicosmo**, 2004. 'Antitumor agents: Taxol and taxane production by yew cell culture. In RA Myers. Encyclopedia of Molecular Cell Biology and Molecular Medicine'. *Wiley-VCH, Weinheim*. pp: 415-438
- Frense, D.**, 2007. 'Taxanes: perspectives for biotechnological production'. *Appl Microbiol Biot* 73:1233–40.
- Gandhi, SG., V. Mahajan and YS. Bedi**, 2015. 'Changing trends in biotechnology of secondary metabolism in medicinal and aromatic plants'. *Planta* 241: 303-317.
- Holmes, F. A., A.P. Kudelka, J.J.Kavanagh, M . H. Huber, J. A. Ajani and V. Valero**, 1995. 'Taxane anticancer agents'. Basic science and current status, (Georg GL, Chen TT, Ojima I, Vyas DM, ed.), PP: 3`-57. *American Chemical Society*, Washington DC.
- Hezari, M., NG. Lewis and R. Croteau**, 1995. 'Purification and characterization of taxa-4(5),11(12)-diene synthase from pacific yew (*Taxus brevifolia*) that catalyzes the first committed step of taxol biosynthesis'. *Archives of Biochemistry and Biophysics* 322: 437-444.
- Hortuwitz, SB**, 1992. 'Mechanism of ction of taxol'. *Trends. Pharmacol. Sci.* 13: 134-136.
- Itokawa, H. and K-H. Lee**, 2002. 'Taxus the genus Taxus'. *Taylor and Francis*. London and new york.
- Itokawa, H. and K. H. Lee**, 2003. 'Taxus: The genus of Taxus'. London: Taylor and Francis.
- Jordon, M.A. and L. Wilson**, 1995. 'Microtubule polymerization dynamics, mitotic block, and cell death by paclitaxel at low concentration'. *American Chemical Society Symposium Series*, Vol. 583, Chapter X 138 – 153.
- Jennewein, S. and R. Croteau**, 2001. 'Taxol: biosynthesis, molecular genetics, and biotechnological applications'. *Applied Microbiology and Biotechnology*, 57, 13-19.
- Jennwein, S. and R. Croteau**, 2001. 'Taxol: biosynthesis, molecular genetics, and biotechnological applications'. *Applied Microbiology and Biotechnology*, 57(2-1): 9-13.
- Jennewein, S. and R. Croteau**, 2001. 'Taxol biosynthesis, molecular genetics and biotechnological applications'. *Applied Microbiology and Biotechnology* 57(1-2): 9-13
- Jha, S., D. Sanyal, B. Ghosh and T. B. Jha**, 1998. 'Improved Taxol yield in cell suspension culture of *Taxus wallichiana* (Himalayan Yew)'. *Planta Medica*, 64, 270-272.
- Ketchum, R. E. B., D. M. Gibson, R. B. Croteau and M. L. Shuler**, 1999. 'The kinetics of taxoid accumulation in cell suspension cultures of *Taxus* following elicitation with methyl jasmonate'. *Biotechnology and Bioengineering*, 62, 97-105.
- Khosroushahi, A. Y., M. Valizadeh, A. Ghasempour, M. Khosrowshahli, H. Naghdibadi, M. R. Dadpour and Y. Omid**, 2006. 'Improved Taxol production by combination of inducing factors in suspension cell culture of *Taxus baccata*'. *Cell Biology International*, 30, 262-269.
- Kajani, A. A., S. Moghim and M. R. Mofid**, 2012. 'Enhanced taxane production and secretion from *Taxus baccata* cell culture by adding dimethyl sulfoxide'. *Biotechnology and Applied Biochemistry*. (wileyonlinelibrary.com).
- Kolewe, ME., V. Gaurav, SC. Roberts**, 2008. 'Pharmaceutically active natural product synthesis and supply via plant cell culture technology'. *Mol Pharmaceut*, 5:243–56.
- Liu, WC., T. Gong and P. Zhu**, 2016. 'Advances in exploring alternative Taxol sources'. *Royal*

- Society of Chemistry Advances*; 6: 48800-48809.
- Lee, D-H., S-G. Kim, S. Mun and JH. Kim**, 2010. 'Evaluation of feeding and mixing conditions for fractional precipitation of paclitaxel from plant cell cultures'. *Process Biochemistry*, 45(7), 1134-1140.
- Linden, J. C. and M. Phisalaphong**, 2000. 'Oligosaccharides potentiate methyl jasmonate induced production of paclitaxel in *Taxus canadensis*'. *Plant Science*, 158, 41-51.
- Mossadegh, A.**, 1993. 'Yew tree'. Research Report of University of California, Berkeley.
- Mihaljevic, S. and I. Bjedow**, 2002. 'Effect of explant source and growth regulators on in vitro callus growth of *Taxus baccata* L. washingtonii'. *Food Technology. Biotechnology*, 40(4): 299-303.
- Malik, S., R. Cusido, M. Mirjalili, E. Moyano, J. Palazon and M. Bonfil**, 2011. 'Production of the anticancer drug taxol in *Taxus baccata* suspension cultures: A review'. *Process Biochem.* 46: 23-34.
- Mustafa, NR., W. DeWinter, F. Vanlren, R. Verpoorte**, 2011. 'Initiation, growth and cryopreservation of plant cell suspension cultures'. *Nature Protoc* 6:715-742
- Mejía-Teniente, L., I. Torres-Pacheco, M.M. González-Chavira, R.V. Ocampo-Velazquez, G. Herrera-Ruiz, A.M. Chapa-Oliver and R.G. Guevara-González**, 2010. 'Use of elicitors as an approach for sustainable agriculture'. *African Journal of Biotechnology*. 9, 9155-9162.
- Nicolau, K. C., Z. Yang, and J. Liu**, 1994. 'Total synthesis of taxol'. *Nature Biotechnology*, 367(64), 630-634.
- Nims, E., C. P. Dubois, S. C. Roberts and R. Walker**, 2006. 'Expression profiling of genes involved in paclitaxel biosynthesis for targeted metabolic engineering'. *Meta-engineering*, 8:385-394.
- Onrubia, M., RM. Cusido, K. Ramirez, L. Hernandez-Vazquez, E. Moyano, M. Bonfill and J. Palazon**, 2013. 'Bioprocessing of plant in vitro systems for the mass production of pharmaceutically important metabolites: Paclitaxel and its derivatives'. *Current Medicinal Chem.* 20: 880-891.
- Roberts, SC. and ML. Shuler**, 1997. 'Large-scale plant cell culture'. *Curr Opin Biotech* 8:154-9.
- Rao, SR., and GA. Ravishankar**, 2002. 'Plant cell cultures: Chemical factories of secondary metabolites'. *Biotechnology Advances* 20: 101-153.
- Shoeb, M.**, 2006. 'Anticancer agents from medicinal plants'. *Bangladesh journal of pharmacology.*;1(2):35-41.
- Siegel, R. and A. Jemal**, 2012. 'Cancer Facts & Figures'. *American Cancer Society*. 404: 1-64.
- Suffness, M. and M.E. Wall**, 1995. 'Discovery and development of taxol'. In: *Taxol: Science and applications*, press, Boca Raton, FL.
- Schippmann, U.** 2001. 'Medicinal plants significant trade study'. Germany: German Federal Agency for Nature Conservation Bonn.
- Stierle, A., G. Strobel, D. Stierle, P. Grothaus and G. Bignami**, 1995. 'The search for a taxol-producing microorganism among the endophytic fungi of the pacific yew, *Taxus brevifolia*'. *Journal of Natural Products*, 58(9), 1315-1324.
- Sabater-Jara, A. B. and M. A. Pedreño**, 2013. 'Use of α -cyclodextrins to enhance phytosterol production in cell suspension cultures of carrot (*Daucus carota* L.)'. *Plant Cell Tissue and Organ Culture*, 114: 249-58.
- Tabata, H.**, 2004. 'Paclitaxel production by plant-cell-culture technology'. *Advances in Biochemical Engineering Biotechnology*, 87, 1-23.
- Vazquez-Hernandez, C., A. Feregrino-Perez, I. Perez-Ramirez, R.V. Ocampo-Velazquez, E. Rico-García, I. Torres-Pacheco and R. G. Guevara-Gonzalez**, 2019. 'Controlled elicitation increases steviol glycosides (SGs) content and gene expression-associated to biosynthesis of SGs in *Stevia rebaudiana* B'. cv. Morita II. *Industrial Crops & Products*, 139: 111479.
- Vargas-Hernandez, M., I. Macias-Bobadilla, R.G. Guevara-Gonzalez, Sd.J. Romero-Gomez, E. Rico-Garcia, R.V. Ocampo-Velazquez, Ld.L. Alvarez-Arquieta, and I. TorresPacheco**, 2017. 'Plant hormesis management with bio stimulants of biotic origin in agriculture'. *Frontiers Plant Sciences*. 8, 1762.
- Wall, M.E.** 1993. 'Campotohein and taxol'. In: Lednicer D., (ed.) *Chronicles of Drug*

Discovery, Washington DC, pp:153-165. American Chemical Society.

Wani, M.C., H.L.Taylor, M.E. wall, P. Coggon and A.T. Mcphail, 1971. 'Plant antitumor agent The isolation and structure of taxol, a novel ant leukemic and antitumor agent from *Taxus berevifolia*'. *Journal of the American Chemical Society*, 93: 2325-2327.

Walker, K. and R. Croteau, 2000. 'Molecular cloning of a 10-deacetylbaaccatin III-10-O-acetyl transferase cDNA from *Taxus* and functional expression in *Escherichia coli*'. *Proceedings of the National Academy of Sciences USA*. 97 (2): 583-587.

Walker, K., K. Klettke, T. Akiyama and RB. Croteau, 2004. 'Cloning, heterologous expression, and characterization of a phenylalanine amino mutase involved in Taxol biosynthesis'. *Journal of Biological Chem.* 279: 53947-54.

Walker, K., S. Fujisaki, R. Long and R. Croteau, 2002. 'Molecular cloning and heterologous expression of the C-13 phenylpropanoid side Chain-CoA acyltransferase that functions in Taxol biosynthesis'. *Proceedings of the National Academy of Sciences USA* 99 (20): 12715-12720.

Walker, K., R. Long and R. Croteau, 2002. 'The final acylation step in Taxol biosynthesis: cloning of the taxoid C13-side-chain N-benzoyltransferase from *Taxus*'. *Proceedings of the National Academy of Sciences USA* 99: 9166-71.

Zhong, J., 2002. 'Plant cell culture for production of paclitaxel and other taxanes'. *Journal of Bioscience and Bioengineering*, 94(6), 591-599.

Zhang, C. H., J. Y. Wu and G. Y. He, 2002. 'Effects of inoculum size and age on biomass growth and paclitaxel production of elicitor-treated *Taxus yunnanensis* cell cultures'. *Applied Microbiology and Biotechnology*, 60, 396-402.

Zhang, C. H. and J. Y. Wu, 2003. 'Ethylene inhibitors enhance elicitor-induced paclitaxel production in suspension cultures of *Taxus* spp. cells. *Enzyme and Microbial Technology*, 32, 71-77.

Zhang, C. H., X. G. Mei, L. Liu and L. J. Yu, 2000. 'Enhanced paclitaxel production induced by the combination of elicitors in cell suspension cultures of *Taxus chinensis*'. *Biotechnology Letters*, 22: 1561-156