

# Automatic Diagnosis of Breast Cancer in Histopathologic Images Based on Convolutional AutoEncoders and Reinforced Feature Selection

A. Abdulhussain Fadhil<sup>1\*</sup>, M. Adnan<sup>2</sup>, H. Radhi<sup>3</sup>, M. Al-Mualm<sup>4</sup>, M. H. Alubaidy<sup>5</sup>, M. Salih<sup>6</sup>, and S. Jaafar Saadoon<sup>7</sup>

1- College of Medical Technology, Medical Lab Techniques, Al-farahidi University, Iraq.

Email: alialshamary412@yahoo.com (Corresponding author)

2- Anesthesia Techniques Department, Al-Mustaqbal University College, Babylon, Iraq

Email: miaad.adnan@uomus.edu.iq

3- College of MLT, University of Ahl Al Bayt, Kerbala, Iraq.

Email: hamza.radhi@gmail.com

4- Department of Medical Laboratories Technology, AL-Nisour University College, Baghdad, Iraq.

Email: mmualm@yahoo.com

5- Al-Hadi University College, Baghdad, 10011 Iraq.

Email: dp.dentistryb@huc.edu.iq

6- Department of pharmacy, Ashur University College, Baghdad, Iraq.

Email: mohamedsalih2008@gmail.com

7- College of Pharmacy, Al-Ayen University, Thi-Qar, Iraq.

Email: sarah.j.s@alayen.edu.iq

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## ABSTRACT:

Breast cancer is one the most ubiquitous types of cancer which affect a considerable number of women around the globe. It is a malignant tumor, whose origin is in the glandular epithelium of the breast and causes serious health-related problems for patients. Although there is no known way of curing this disease, early detection of it can be very fruitful in terms of reducing the negative ramifications. Thus, accurate diagnosis of breast cancer based on automatic approaches is demanded immediately. Computer vision-based techniques in the analysis of medical images, especially histopathological images, have proved to be extremely performant. In this paper, we propose a novel approach for classifying malignant or non-malignant images. Our approach is based on the latent space embeddings learned by convolutional autoencoders. This network takes a histopathological image and learns to reconstruct it and by compressing the input into the latent space, we can obtain a compressed representation of the input. These embeddings are fed to a reinforcement learning-based feature selection module which extracts the best features for distinguishing the normal from the malicious images. We have evaluated our approach on a well-known dataset, named BreakHis, and used the K-Fold Cross Validation technique to obtain more reliable results. The accuracy, achieved by the proposed model, is 96.8% which exhibits great performance.

**KEYWORDS:** Breast Cancer Detection, Convolutional Autoencoders, Feature Selection, Reinforcement Learning, Histopathology

## 1. INTRODUCTION

Cancer is a group of illnesses in which the body's cells congregate to create lumps known as malignant tumors [1]. These cells proliferate across the surrounding tissues, expand uncontrollably, and suffocate the healthy cells [2]. From ancient times to the present, cancer has been one of the most serious illnesses to endanger human health. According to a study done in

2018, there will be an estimated 18.1 million new cases of cancer added to the projected 9.6 million cancer cases already present in the world [3].

The prevalence of breast cancer (BC) is rising progressively in both industrialized and developing nations (considered the second most common cancer among women). According to the Nottingham score, the evaluation of nuclear pleomorphism, tubule

development, and the mitotic count is used to grade BC [4]. Epithelial cell nuclei in a healthy breast are homogeneous in size and shape [5]. Malignant epithelial cell nuclei, however, take on a non-uniform, darker, and bigger shape. The term "nuclear pleomorphism" refers to this change. The percentage of cancerous cells that form tubules in a typical duct structure is represented [6]. One of the most crucial proliferation parameters, the mitotic count, provides crucial diagnostic data needed for BC histological grading [7].

Using image processing and machine learning approaches, quick tumor identification and diagnosis may now significantly improve the accuracy of a BC diagnosis [8]. Clinical illness diagnosis, treatment evaluation, and the detection of problems in several human organs, including the eye, lungs, brain, breast, and stomach, all depend heavily on medical imaging [9]. Medical imaging is a variety of methods used to examine the human body in order to identify, monitor, or cure diseases [10].

The histopathologic diagnosis continues to be the gold standard for cancer diagnosis despite significant advances in medical science [11]. Images taken under a microscope of the tissues used to study illness are known as histopathological images [12]. Because of the nature of histological pictures and the sharp increase in labor, this task takes a long time, and the results could be influenced by the pathologist's subjective judgment.

Hitherto, a variety of machine learning and deep learning-based algorithms have been put forward for detecting malignant samples in histopathologic images [13]. In [14], Spanhol et al. trained a distinct version of the AlexNet CNN model using a collection of pixel components from HPis. These pieces, which are 32\*32 and 64\*64 in size, were created utilizing sliding window and randomization techniques. According to the magnification factor, the best accuracy for binary categorization (malignant and benign) was between 80 and 90 percent. Additionally, two distinct CNN models were created by Bayramoglu et al. [15] to forecast two classes (binary classification). While multi-classification accuracy fluctuated between 80 and 83 percent, binary classification accuracy fluctuated between 82 and 85 percent. Furthermore, Wei et al. [16] introduced a CNN model for binary classification called BiCNN that has three convolutional layers and three pooling layers. The BiCNN model's performance was contrasted with that of pre-trained CNN models like VGGNet and AlexNet. Pre-trained CNN models were surpassed by the BiCNN model, which had accuracy levels between 97.56 and 97.97 percent. In addition, CNN and Bidirectional Long Short Term Memory (BiLSTM) models were utilized for binary classification by Budak et al. [17]. Both the CNN model and the BiLSTM model underwent independent training for the tasks of deep feature extraction and classification. For

various magnification factors, this hybrid model achieved accuracy scores between 93.61 percent and 96.32 percent. Li et al. [18] introduced a deep learning model for breast cancer diagnosis that is based on the CNN model and uses an end-to-end learning procedure. With this model, the categorization accuracy was 90.0 percent. To improve classification performance, Thuy et al. [19] suggested a hybrid deep learning model including VGG16 and VGG19 CNN models and a generative adversarial network (GAN). On the 2-class BreakHis dataset, this technique has a classification accuracy of 98.1 percent.

In this paper, we propose an approach based on Convolutional AutoEncoders (CAEs) and reinforcement learning-based feature selection for diagnosing breast cancer from histopathological images. Our approach does not need any special preprocessing step except for normalization. In addition to this, a dataset collected from real subjects in hospitals has been gathered to train and evaluate the proposed pipeline. Overall, in this work, our contributions are fourfold as follows:

1. A robust approach for diagnosing breast cancer is proposed in which the power of CAEs is utilized.
2. A feature selection module based on reinforcement learning is integrated with the object of optimizing the bottleneck feature vector generated by the CAE.
3. A dataset of 60000 samples has been collected from real hospitals and this adds to the reliability of the results.
4. Our proposed approach achieves competitive results, compared with the previous works.

The rest of the paper is outlined as follows: Section 2 explicates the details of the proposed methodology. Section 3 contains our results and experimental setup. Section 4 includes the conclusion.

## 2. MATERIALS AND METHODS

### 2.1. Overview

An overview of the proposed methodology is depicted in Fig. 1. As is seen in Fig. 1, the input batch of images, after being preprocessed, is fed into the model and the model learns to reconstruct the original image. This way, the bottleneck vector is a representation that can be used to be classified. Our proposed method contains a feature selection stage, where we utilize the power of RL to select the best elements in the feature vector.

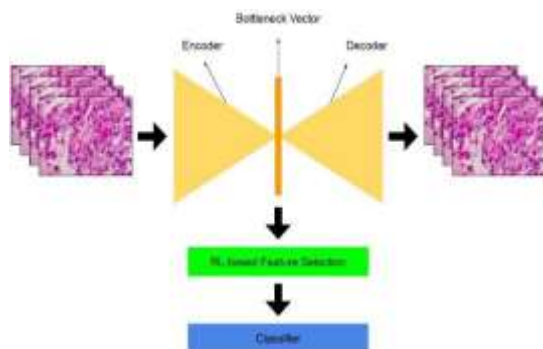


Fig. 1. The overview of the proposed methodology.

## 2.2. Dataset

In this study, we have used a dataset containing 60000 samples in two classes, namely normal and cancerous. The samples are collected from three well-known health centers in Iraq, namely Saint Raphael (Al Rahibat) Hospital and Ibn Al-Bitar Hospital. Fig. 2 demonstrates some samples from the dataset from the two classes.

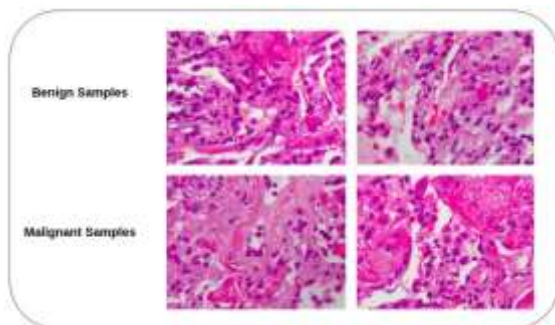


Fig. 2. The samples from both classes in the dataset.

Using specialized cameras and a microscope, together with an automated computerized process, it is possible to acquire histopathology pictures [20]. The biopsy specimen is embedded in wax and stained with one or more dyes in order to examine the different architecture and components of tissues under a microscope [21]. Pathologists utilize staining techniques to separate cellular components for structural as well as component viewing of tissue for diagnosis [22]. There are five steps in it, and each one has the potential to impact the final image's quality. Fixing: Chemical fixation is used to preserve biological tissue samples. There are several methods of fixation, but formaldehyde or glutaraldehyde solution fixation is the method that is most frequently used in the biomedical area to safeguard the cells. To avoid tissue autolysis and putrefaction, this crucial stage is tissue preparation; Processing: Dehydration and clearing are the two key steps in tissue processing, which is a vital stage. Dehydration is used to remove water from the viscous tissue and replace it with an alcohol solution that hardens it. This procedure aids

in cutting extremely thin slices of the material. Clearing entails removing the dehydrator using a substance that serves as both the embedding paraffin and the dehydrating agent's solvent. This procedure is crucial because perfect microscopic analysis requires the correct orientation of the tissue; Sectioning: This procedure is necessary to produce ultra-thin slices of tissue samples that are sufficient to enable clear observation of the microstructure characterization of the cells using microscopy techniques. Afterward, transfer the ultra-thin slices of the sample on a fresh glass slide; Staining: Staining the tissue and mounting it on the slide are the last steps in preparing it for light microscopy. Staining enhances the tissue's contrasts and draws attention to some particular characteristics that would otherwise be virtually unnoticeable under the microscope. Although there are many other types of stains, H & E staining is the most used form for histology [23].

## 2.3. Convolutional AutoEncoders

Due to the remarkable performance of convolutional neural networks, researchers have proposed a variety of such models in various fields [24]-[25]-[26]-[27]-[28]. A particular kind of feedforward convolutional neural network called an autoencoder uses input and output to be identical [29]. They reduce the input's dimension before using this representation to recreate the output. The code, also known as the latent-space representation, is an efficient "summary" or "compression" of the input [30]. Encoder, code, and decoder are the three parts of an autoencoder. The input is compressed by the encoder, which also creates a code. The decoder then reconstructs the input exclusively using the code [31]. Convolutional AutoEncoders (CAEs) use the convolution operator to exploit this observation [32]. Rather than manually engineer convolutional filters, we let the model learn the optimal filters that minimize the reconstruction error [33]. These filters can then be used in any computer vision task. CAEs are the state of art tools for unsupervised learning of convolutional filters. Once these filters have been learned, they can be applied to any input to extract features [34]. These features can be used to do any task that requires a compact representation of the input, like classification [35]. The primary distinction between the conventional interpretation of CNN and CAE is the former's end-to-end training in the acquisition of filters and the combining of features with the goal of categorizing input [36]. The latter are merely taught filters that can extract information from the input and be used to rebuild it. Table 1 demonstrates the train, validation, and test distribution for the dataset.

**Table 1.** The distribution of train/validation and test sets.

	Train	Validation	Test
Malignant	19200	4800	6000
Benign	19200	4800	6000
Number of samples	38400	9600	12000

#### 2.4. Feature Selection using Reinforcement Learning

The range of variables or characteristics that may be utilized to describe a specific predictor of interest keeps expanding exponentially as the cost of data gathering falls [37]. Therefore, the key to properly training a machine learning model is to select the most distinctive characteristics that decreases variance without endangering the bias of our models [38]. Finding these traits is also essential for optimal computing cost, predictability, and interpretability [39]. While statistical techniques like shrinkage, subset selection, and dimensionality reduction have been used to choose the best set of features, some other approaches in the literature have treated the task of feature selection as a search problem where each state in the search space is a potential feature subset [40].

Inspired by [41], using the Reinforcement Learning technique, where the state space consists of all conceivable subsets of the features and action is any feature that is included in the model, we attempt to address the feature selection problem. The amount of features that are missing from the model affects the action space for each state [42]. By doing so, we may avoid having an excessively sparse state space, decrease the search space for the next optimal action, and speed up computation [43]. The scoring accuracy of the machine learning algorithm used to assess the prediction strength of the present state determines the reward function [44]. The reward is defined as equation 1:

$$R_f = Accuracy_{t+1} - Accuracy_t \quad (1)$$

Based on the reward received and the value of the state when it was previously visited, we applied the Temporal Difference (TD) method to evaluate the state's worth [45]. The formulation of this issue allows it to work with a high-dimensional feature space and is strong enough to handle any non-linear relationships between the predictors and the response variable. For the purpose of determining the state value for each chosen subset, we employed the Support Vector Machines (SVM) classifier [46]. SVM exhibits strong behavior across a wide range of learning tasks. Additionally, they are completely automated, therefore classifier parameter adjustment is not necessary. SVM classifiers also work well in high-dimensional spaces and are particularly

resilient to difficulties with non-linear classification [47].

### 3. EXPERIMENTAL RESULTS

#### 3.1. Experimental Setup

This subsection includes all the mediums used in implementing the proposed methodology. We used Python 3.10 as the programming language and Pytorch 1.10 as the deep learning framework. The Central Processing Unit (CPU) of the machine we used for training is core i7, 3.8 GHz and its Graphical Processing Unit (GPU) is GeForce RTX 1050. Additionally, the learning rate for training the autoencoder is 0.004 with a batch size of 64. The model is trained for 55 epochs and the optimizer used in the implementation is Adam.

Furthermore, Table 2 shows the architecture of the CAE used in this study. Table 2 contains both the architecture convolutional encoder of the model and its decoder part.

**Table 2.** The CAE that was used in this study.

Network	Layer Type	Parameters	Non-Linear Activation Function
Input	Input	480	-
Encoder	Conv	64	ReLU
	Pooling	32	-
	Conv	16	ReLU
	Pooling	8	-
Decoder	Conv	8	ReLU
	Upsampling	16	-
	Conv	32	ReLU
	Upsampling	64	-

#### 3.2. Classification metrics

We have used the metrics introduced in Table 3. for evaluating our proposed approach. These metrics are used in order to prove the efficacy of a classifier in machine learning.

Fig. 3 demonstrates a Confusion Matrix (CM), which includes 4 important entities named True Positive (TP), True Negative (TN), False Positive (FP), and False Negatives (FN). TP is the number of malignant samples that are classified correctly. TN is the number of benign samples that are classified correctly. FP is the number of samples that are actually benign but classified as malignant and FN is the number of benign samples that are classified as malignant.

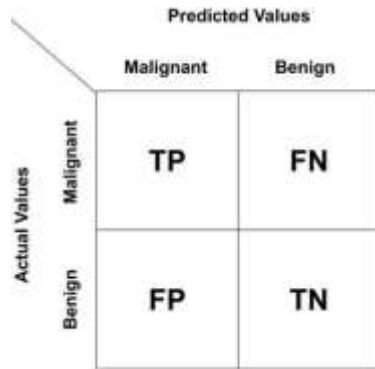


Fig. 3. A Confusion Matrix (CM) that is used for evaluating classifiers

Table 3. The metrics used for evaluating our proposed method.

Metric	Calculation
Accuracy	$\frac{TP + TN}{TP + FP + FN + TN}$
Precision	$\frac{TP}{TP + FP}$
Recall	$\frac{TP}{TP + FN}$
F1-Score	$\frac{2 \times Precision \times Recall}{Precision + Recall}$

3.3. Classification results

This section includes the results achieved by the proposed classifier based on the metrics introduced in section 3.2. Fig. 4 and Table 4 detail the CM and the results obtained by our proposed algorithm.

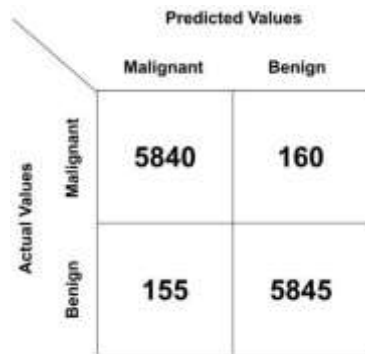


Fig. 4. Training and validation loss vs. epoch curve.

Table 4. Results achieved by the proposed methodology.

Metric	Accuracy (%)	Recall (%)	Precision (%)	F1-Score (%)
Obtained Value	97.38	97.32	97.41	97.35

Moreover, Fig. 5 illustrates the loss vs. epoch curve for training the autoencoder. Fig. 6 demonstrates the

reconstructed images using the autoencoder for two samples.

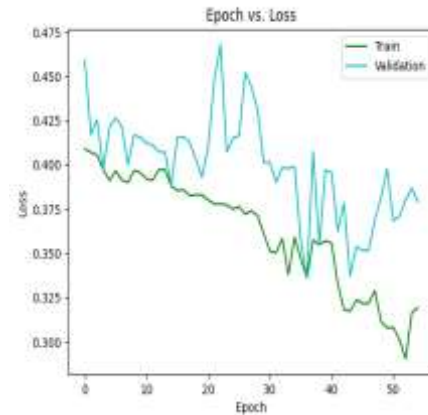


Fig. 5. Training and validation loss vs. epoch curve.

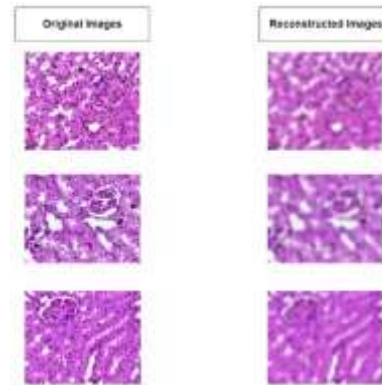


Fig. 6. Samples of original and reconstructed images using the proposed CAE.

Furthermore, Table 5 details comparison between our proposed methodology and other state-of-the-art research works. This comparison is done using the accuracy achieved by each methodology.

Table 5. Comparison between our proposed methodology and other research works

Research	Accuracy (%)
[14]	90.00
[15]	85.00
[16]	97.97
[17]	96.32
[18]	90.00
[19]	98.10
Our proposed method	97.38

Based on Table 5, the accuracy obtained by our proposed methodology is thoroughly competitive with the other works. This shows the reliable performance of the algorithm for classifying histopathological images.

Moreover, based on the F1-Score that is achieved by our proposed method, we can claim that the model has significant performance in recognizing both TP and TN samples within the dataset used in this study.

#### 4. CONCLUSION

In this study, we have proposed an algorithm for atomizing the diagnosis procedure for breast cancer images. Our algorithm is able to analyze histopathological images without the demand for any special preprocessing step. Based on our extensive experiments, the performance of the model can be claimed to be remarkably good. Since our experiments have been conducted on a large dataset gathered from real hospitals, it can be argued that the proposed pipeline can be utilized efficiently in clinical contexts with ease.

#### REFERENCES

- [1] Goodall GJ, Wickramasinghe VO. RNA in cancer. *Nature Reviews Cancer*. 2021 Jan;21(1), pp. 22-36.
- [2] Waks AG, Winer EP, “**Breast cancer treatment: a review**,” *Jama*. 2019 Jan 22;321(3), pp. 288-300.
- [3] Ferlay J, Colombet M, Soerjomataram I, Parkin DM, Piñeros M, Znaor A, Bray F, “**Cancer statistics for the year 2020: An overview**,” *International journal of cancer*. 2021 Aug 15;149(4), pp.778-89.
- [4] Elsharawy KA, Gerds TA, Rakha EA, Dalton LW, “**Artificial intelligence grading of breast cancer: a promising method to refine prognostic classification for management precision**,” *Histopathology*. 2021 Aug;79(2), pp.187-99.
- [5] Singh AV, Maharjan RS, Kanase A, Siewert K, Rosenkranz D, Singh R, Laux P, Luch A, “**Machine-learning-based approach to decode the influence of nanomaterial properties on their interaction with cells**,” *ACS Applied Materials & Interfaces*. 2020 Dec 29;13(1), pp. 1943-55.
- [6] Sohail A, Khan A, Wahab N, Zameer A, Khan S, “**A multi-phase deep CNN based mitosis detection framework for breast cancer histopathological images**,” *Scientific Reports*. 2021 Mar 18;11(1), pp. 1-8.
- [7] Budak Ü, Cömert Z, Rashid ZN, Şengür A, Çıbuk M, “**Computer-aided diagnosis system combining FCN and Bi-LSTM model for efficient breast cancer detection from histopathological images**,” *Applied Soft Computing*. 2019 Dec 1;85:105765.
- [8] Saxena S, Gyanchandani M, “**Machine learning methods for computer-aided breast cancer diagnosis using histopathology: a narrative review**,” *Journal of medical imaging and radiation sciences*. 2020 Mar 1;51(1), pp. 182-93.
- [9] Suganyadevi S, Seethalakshmi V, Balasamy K, “**A review on deep learning in medical image analysis**,” *International Journal of Multimedia Information Retrieval*. 2022 Mar;11(1), pp. 19-38.
- [10] Zhang J, Xie Y, Wu Q, Xia Y, “**Medical image classification using synergic deep learning**,” *Medical image analysis*. 2019 May 1;54, pp. 10-9.
- [11] Kiani A, Uyumazturk B, Rajpurkar P, Wang A, Gao R, Jones E, Yu Y, Langlotz CP, Ball RL, Montine TJ, Martin BA, “**Impact of a deep learning assistant on the histopathologic classification of liver cancer**,” *NPJ digital medicine*. 2020 Feb 26;3(1), pp. 1-8.
- [12] Zhai J, Shen W, Singh I, Wanyama T, Gao Z, “**A review of the evolution of deep learning architectures and comparison of their performances for histopathologic cancer detection**,” *Procedia Manufacturing*. 2020 Jan 1;46, pp. 683-9.
- [13] Houssein EH, Emam MM, Ali AA, Suganthan PN, “**Deep and machine learning techniques for medical imaging-based breast cancer: A comprehensive review**,” *Expert Systems with Applications*. 2021 Apr 1;167:114161.
- [14] Bejnordi BE, Veta M, Van Diest PJ, Van Ginneken B, Karssemeijer N, Litjens G, Van Der Laak JA, Hermsen M, Manson QF, Balkenhol M, Geessink O, “**Diagnostic assessment of deep learning algorithms for detection of lymph node metastases in women with breast cancer**,” *Jama*. 2017 Dec 12;318(22), pp. 2199-210.
- [15] Thuy MB, Hoang VT, “**Fusing of deep learning, transfer learning and gan for breast cancer histopathological image classification**,” *In International Conference on Computer Science, Applied Mathematics and Applications* 2019 Dec 19, pp. 255-266, Springer, Cham.
- [16] Bardou D, Zhang K, Ahmad SM, “**Classification of breast cancer based on histology images using convolutional neural networks**,” *IEEE Access*. 2018 May 1;6:24680-93.
- [17] Boumaraf S, Liu X, Wan Y, Zheng Z, Ferkous C, Ma X, Li Z, Bardou D, “**Conventional machine learning versus deep learning for magnification dependent histopathological breast cancer image classification: A comparative study with visual explanation**,” *Diagnostics*. 2021 Mar 16;11(3):528.
- [18] Song Y, Zou JJ, Chang H, Cai W, “**Adapting fisher vectors for histopathology image classification**,” *In 2017 IEEE 14th international symposium on biomedical imaging (ISBI 2017)* 2017 Apr 18, pp. 600-603. IEEE.
- [19] Zhi W, Yueng HW, Chen Z, Zandavi SM, Lu Z, Chung YY, “**Using transfer learning with convolutional neural networks to diagnose breast cancer from histopathological images**,” *In International Conference on Neural Information Processing 2017 Nov 14*, pp. 669-676, Springer, Cham.
- [20] Zhan X, Cheng J, Huang Z, Han Z, Helm B, Liu X, Zhang J, Wang TF, Ni D, Huang K, “**Correlation analysis of histopathology and proteogenomics data for breast cancer**,” *Molecular & Cellular Proteomics*. 2019 Aug 9;18(8):S37-51.
- [21] Li X, Shen X, Zhou Y, Wang X, Li TQ, “**Classification of breast cancer histopathological images using interleaved DenseNet with SENet (IDSNet)**,” *PloS one*. 2020 May 4;15(5):e0232127.
- [22] Al-Haija QA, Adebajo A, “**Breast cancer diagnosis in histopathological images using ResNet-50 convolutional neural network**,” *In 2020 IEEE International IOT, Electronics and Mechatronics Conference (IEMTRONICS)* 2020 Sep 9, pp. 1-7, IEEE.

- [23] Parvin F, Hasan MA, “A comparative study of different types of convolutional neural networks for breast cancer histopathological image classification,” *In 2020 IEEE Region 10 Symposium (TENSYP)* 2020 Jun 5, pp. 945-948, IEEE.
- [24] Sharghi E, Nourani V, Najafi H, Molajou A, “Emotional ANN (EANN) and wavelet-ANN (WANN) approaches for Markovian and seasonal based modeling of rainfall-runoff process,” *Water resources management*. 2018 Aug;32(10), pp. 3441-56.
- [25] Nourani V, Razzaghzadeh Z, Baghanam AH, Molajou A, “ANN-based statistical downscaling of climatic parameters using decision tree predictor screening method. Theoretical and Applied Climatology,” 2019 Aug;137(3), pp. 1729-46.
- [26] Mehr AD, Nourani V, Hrnjica B, Molajou A, “A binary genetic programming model for teleconnection identification between global sea surface temperature and local maximum monthly rainfall events,” *Journal of Hydrology*. 2017 Dec 1;555, pp. 397-406.
- [27] Wang J, Zhu H, Wang SH, Zhang YD, “A review of deep learning on medical image analysis. Mobile Networks and Applications,” 2021 Feb;26(1), pp. 351-80.
- [28] Adegun AA, Viriri S, Ogundokun RO, “Deep learning approach for medical image analysis,” *Computational Intelligence and Neuroscience*. 2021 May 8;2021.
- [29] Ahmed AS, El-Behaidy WH, Youssif AA, “Medical image denoising system based on stacked convolutional autoencoder for enhancing 2-dimensional gel electrophoresis noise reduction,” *Biomedical Signal Processing and Control*. 2021 Aug 1;69:102842.
- [30] Nogales A, Garcia-Tejedor AJ, Monge D, Vara JS, Antón C, “A survey of deep learning models in medical therapeutic areas,” *Artificial Intelligence in Medicine*. 2021 Feb 1;112:102020.
- [31] Mutabazi E, Ni J, Tang G, Cao W, “A review on medical textual question answering systems based on deep learning approaches,” *Applied Sciences*. 2021 Jan;11(12), pp. 5456.
- [32] Dong S, Wang P, Abbas K, “A survey on deep learning and its applications,” *Computer Science Review*. 2021 May 1;40:100379.
- [33] Souravlas S, Anastasiadou S, Katsavounis S, “A Survey on the Recent Advances of Deep Community Detection,” *Applied Sciences*. 2021 Aug 4;11(16):7179.
- [34] Hammouche R, Attia A, Akhrouf S, Akhtar Z, “Gabor filter bank with deep autoencoder based face recognition system,” *Expert Systems with Applications*. 2022 Feb 26:116743.
- [35] Ghojogh B, Ghodsi A, Karray F, Crowley M, “Generative adversarial networks and adversarial autoencoders: Tutorial and survey,” *arXiv preprint arXiv:2111.13282*. 2021 Nov 26.
- [36] Baur C, Denner S, Wiestler B, Navab N, Albarqouni S, “Autoencoders for unsupervised anomaly segmentation in brain MR images: a comparative study,” *Medical Image Analysis*. 2021 Apr 1;69:101952.
- [37] Liu K, Fu Y, Wu L, Li X, Aggarwal C, Xiong H. “Automated feature selection: A reinforcement learning perspective,” *IEEE Transactions on Knowledge and Data Engineering*. 2021 Sep 24.
- [38] Feng J, Li D, Gu J, Cao X, Shang R, Zhang X, Jiao L, “Deep reinforcement learning for semisupervised hyperspectral band selection,” *IEEE Transactions on Geoscience and Remote Sensing*. 2021 Feb 19;60, pp. 1-9.
- [39] Le N, Rathour VS, Yamazaki K, Luu K, Savvides M, “Deep reinforcement learning in computer vision: a comprehensive survey,” *Artificial Intelligence Review*. 2021 Sep 29, pp. 1-87.
- [40] Kaur A, Guleria K, Trivedi NK, “Feature selection in machine learning: methods and comparison,” *In 2021 International Conference on Advance Computing and Innovative Technologies in Engineering (ICACITE)* 2021 Mar 4, pp. 789-795, IEEE.
- [41] Rasoul S, Adewole S, Akakpo A, “Feature selection using reinforcement learning,” *arXiv preprint arXiv:2101.09460*. 2021 Jan 23.
- [42] Behura A, “The cluster analysis and feature selection: Perspective of machine learning and image processing. Data Analytics in Bioinformatics: A Machine Learning Perspective,” 2021 Feb 1:249-80.
- [43] Obite F, Usman AD, Okafor E, “An overview of deep reinforcement learning for spectrum sensing in cognitive radio networks,” *Digital Signal Processing*. 2021 Jun 1;113:103014.
- [44] Ebrahimi-Khusfi Z, Nafarzadegan AR, Dargahian F, “Predicting the number of dusty days around the desert wetlands in southeastern Iran using feature selection and machine learning techniques,” *Ecological Indicators*. 2021 Jun 1;125:107499.
- [45] Pateria S, Subagdja B, Tan AH, Quek C, “Hierarchical reinforcement learning: A comprehensive survey,” *ACM Computing Surveys (CSUR)*. 2021 Jun 5;54(5), pp. 1-35.
- [46] Chakraborty A, Mitra S, De D, Pal AJ, Ghaemi F, Ahmadian A, Ferrara M, “Determining Protein-Protein Interaction Using Support Vector Machine: A Review,” *IEEE Access*. 2021 Jan 12;9, pp. 12473-90.
- [47] Jueyendah S, Lezgy-Nazargah M, Eskandari-Naddaf H, Emamian SA, “Predicting the mechanical properties of cement mortar using the support vector machine approach,” *Construction and Building Materials*. 2021 Jul 12;291:123396.